

NOVEL CYTOTOXIC DITERPENES FROM THE STEM OF DYSOXYLUM KUSKUSENSE¹

Toshihiro Fujioka, Miyako Yamamoto, Yoshiki Kashiwada, Hiroko Fujii, Kunihide Mihashi, Xashiwada, Hiroko Fujii, Mihashi, Xashiwada, Hiroko Fujii, Hiroko Fujii, Kunihide Mihashi, Xashiwada, Hiroko Fujii, Xa

^aFaculty of Pharmaceutical Sciences, Fukuoka University, Fukuoka 814-0180, Japan

^bNiigata College of Pharmacy, Kamishin'ei-cho 5-13-2, Niigata 950-2081, Japan

^cNatural Products Research Center, Kaohsiung Medical College, Kaoshiung, Taiwan

^dNatural Products Laboratory, Division of Medicinal Chemistry and Natural Products, School of Pharmacy

University of North Carolina, Chapel Hill, North Carolina 27599, U.S.A

Received 29 July 1998; accepted 21 October 1998

Abstract: Three novel diterpenes, dysokusones A (1), B (2), and C (3), were isolated from the stem of *Dysoxylum kuskusense* as cytotoxic substances. The structures were established by spectroscopic examinations. Compounds 1, 2, and 3 were cytotoxic toward HL-60(TB) cells with EC₅₀ values of 2.25, 6.35, and 2.37 μ M, respectively. Compound 1 also displayed cytotoxicity against K-562 and NCI-H522 cells with EC₅₀ values of 5.04 and 4.80 μ M, respectively. © 1998 Elsevier Science Ltd. All rights reserved.

Our previous papers reported the isolation and characterization of 17 triterpene glucosides, cumingianosides A-Q, as well as a trisnor- and a tetranor-triterpene glucoside, cumindysosides A and B, respectively, from a cytotoxic fraction of the leaves of *Dysoxylum cumingianum* (Meliaceae). Among them, cumingianosides A and C exhibited potent selective cytotoxicity against MOLT-4 human leukemia cells with EC₅₀ values of <0.00625 and <0.0045 µM, respectively. In our continuing investigation of cytotoxic compounds in *Dysoxylum* spp. plants as part of our search for novel plant-derived cytotoxic agents, we investigated the cytotoxic constituents in a MeOH extract of the stem of *Dysoxylum kuskusense* (Meliaceae). Subsequent fractionation of the hexane-soluble fraction (20.7 g from 5.4 kg dried stem) by using chromatography on silica gel, MCI-gel CHP 20P, and YMC ODS-A, and by semi-preparative scale HPLC on YMC ODS and Wakopack Sil has led to the isolation of three new diterpenes, named dysokusones A (1, 15 mg), B (2, 17 mg), and C (3, 130 mg), as cytotoxic compounds.

Dysokusone A (1)³ was obtained as a yellow syrup and had the molecular formula $C_{20}H_{32}O_2$. The ¹H nmr spectrum showed one tertiary methyl group (δ 0.85, s), three secondary methyl groups [δ 0.89 (3H, d, J= 6.5 Hz), 0.92 (3H, d, J= 6.5 Hz), and 0.93 (3H, d, J= 6.5 Hz)] and a vinylic methyl group (δ 1.90, t, J= 1.5 Hz). The ¹H and ¹³C nmr spectra also showed a trisubstituted double bond [δ 5.87 (br s); δ 126.42 (d) and 162.91 (s)] and two carbonyl groups (δ 199.11 and 210.56). Examination of the ¹H-¹H COSY and ¹H-¹³C COSY spectra provided two fragment structures shown by bold lines in Figure 1. The long-range couplings in the HMBC spectrum established the connectivity of these fragment units, including methyl groups, carbonyl

groups, and a double bond, as shown in Figure 1, to furnish a prenyleudesmane-type skeleton.

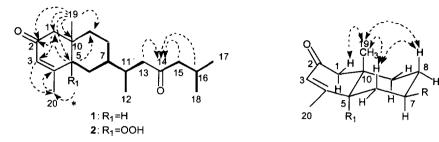


Figure 1. Long-range ¹H-¹³C Correlations (H→C) in 1 and 2 (* Observed in 1)

Figure 2. NOE Correlations in 1 $(R_1 = H)$ and 2 $(R_1 = OOH)$

In the ¹H NMR spectrum of 1, the large coupling constants of H-5, -6, -7, -8 and -9 ($J_{5,6ax} = J_{6ax,7} = J_{7,8ax} = J_{8ax,9ax} = 11.5 \text{ Hz}$) indicated that the B ring is in a chair conformation. NOE between H-1 (δ 2.25) and H-19, H-6ax and H-8ax, H-6ax and H-19, and H-8ax and H-19 together with long-range ¹H-¹H coupling between H-1 (δ 2.17) and H-19 (W arrangement), indicated a *trans* ring fusion in 1.⁴ The chemical shift (δ 0.85) for the C-19 methyl group was also consistent with a *trans* ring fusion. ⁵ In addition, the CD curve of 1 ([θ]₂₄₀ -8307, [θ]₃₃₀ +884) was superimposable on that of α -rotunol (4).⁴ From the observations described above, the structure of dysokusone A was concluded to be represented by formula 1. However, because the small sample size precluded further examination, the absolute configuration of C-11 is still not clear.

Dysokusone B (2)⁶ had the molecular formula $C_{20}H_{32}O_4$, which was 32 units (O_2) more than that of 1. The ¹H and ¹³C NMR spectra of 2 correlated with those of 1 and showed the presence of the same functional groups seen in 1, including one tertiary methyl (δ 1.02, s) three secondary [δ 0.89 (3H, d, J = 5.0 Hz), 0.90 (3H, d, J = 5.0 Hz), and 0.93 (3H, d, J = 6.0 Hz)], and one vinylic methyl (δ 2.05, d, J = 1.5 Hz) groups, a trisubstituted double bond [δ 5.80 (br s); δ 129.27 (d) and 162.88 (s)], and two carbonyl groups (δ 198.65 and 210.85). In addition, an oxygenated quaternary carbon resonance at δ 84.23 and a characteristic one-proton singlet at δ 10.29, combined with the loss of H_2O_2 from the molecular ion in the mass spectrum, indicated the presence of a hydroperoxide group in 2.⁷ The location of the hydroperoxide group was determined to be at C-5 by extensive spectroscopic examinations, including ¹H-¹H COSY and ¹H-¹³C COSY spectra. The *trans* ring fusion of 2 was indicated by the NOE between H-19 and H-1 (δ 1.85) and H-6ax as well as the long-range ¹H-¹H coupling between H-1 (δ 2.55) and H-19 (W arrangement), along with the large coupling constants of H-6, -7, -8, and -9 ($J_{6ax,7} = J_{7,8ax} = J_{8ax,9ax} = 13.5$ Hz), which were similar to those found in 1. This conformation (Fig.

2) was in good agreement with the lowfield shift of the protons located on the same side as the hydroperoxide group^{7b,8} [H-1 α (δ 2.55), H-9 α (δ 1.82), and H-7 (δ 1.90)]. Furthermore, the CD curve of **2** ([θ]₂₃₉ -17560, [θ]₃₂₂ +2026) was similar to that found in **1**, and thus the structure of dysokusone B is represented by formula **2**. The absolute configuration of C-11 is still not clear, again because of the small sample size.

Dysokusone C (3)⁹ contained the same molecular formula ($C_{20}H_{32}O_4$) as 2. The ¹H and ¹³C NMR spectra revealed the same functional groups in 3 as those found in 2. Examination of the ¹H-¹H COSY, ¹H-¹³C COSY, and HMBC spectra indicated that 3 had the same general molecular structure but a different stereostructure from those of 2. In the ¹H NMR spectra of 3, the large coupling constants of H-6, -7, -8, and -9 ($J_{6ax,7} = J_{7,8ax} = J_{8ax,9ax} = 12$ Hz) indicated that the B ring is in a chair conformation. NOE was observed between H-19 and H₂-1, H-20 and H-7, and H-20 and H-6eq in the phase-sensitive NOESY of 3, indicative of a cis ring fusion in 3. The chemical shift (δ 1.23) for the C-19 methyl group, which is typical of cis fused eudesmanes, ⁵ provided additional support for this assignment. This conformation was also consistent with the lowfield shift (δ 3.04) of the H-1 β located on the same side as the hydroperoxide group. ^{7b,8} Furthermore, the CD curve of 3 showed a positive Cotton effect ([θ]₂₃₉ -26483, [θ]₃₂₈ +3641), which was opposite to that reported for the cis-fused eudesmane-type sesquiterpene, β -rotunol (5), ⁴ thus indicating that 3 is represented by formula 3. The configuration of C-11 in 3 was assigned to be S based upon the observation of NOE between H-11 and H-8 as well as between H-12 and H-6eq, though the chirality at C-11 has not been fully investigated at this time, because of insufficient amount of sample.

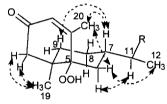


Figure 3. NOE Correlations in 3

The cytotoxicities of dysokusones A (1), B (2), and C (3) against a panel of about 60 tumor cell lines 10 were evaluated at the National Cancer Institute. Dysokusones B (2) and C (3) demonstrated selective cytotoxicity against HL-60(TB) leukemia tumor cells with EC₅₀ values of 6.35 and 2.37 μ M, respectively. Dysokusone A (1) also displayed significant cytotoxicity against the HL-60(TB) cell line with an EC₅₀ value of 2.25 μ M, and showed marginal cytotoxicity against K-562 (leukemia) and NCI-H522 (lung tumor) cells with EC₅₀ values of 5.04 and 4.80 μ M, respectively.

Acknowledgments: The authors wish to thank Dr. A. Mauger, Drug Synthesis and Chemistry Branch, National Cancer Institute, and Dr. B. L. Roberson, Starks C. P., for performing in vitro human tumor cell line assays. They are also indebted to Mr. H. Hanazono for measurement of mass spectra. This investigation was supported by a grant (CA 17625) from the National Cancer Institute awarded to K. H. Lee, and in part by the Ministry of Education, Science, Sports, and Culture of Japan.

	1 ^a		26		3 ^a	
	Н	C	H	C	H	C
1	2.17 (br d, 16.5)	54.48	1.85 (br d, 18)	49.19	2.07 (d, 16)	50.65
	2.25 (br d, 16.5)		2.55 (br d, 18)		3.04 (d, 16)	
2		199.11		198.65		199.59
3	5.87 (br s)	126.42	5.80 (br s)	129.27	6.10 (br s)	132.14
4		162.91		162.88		155.08
5	2.30 (br d, 11.5)	47.87		84.23		85.14
6	1.07 (q, 11.5)	26.00	1.38 (dd, 13.5,14)	27.98	1.33 (t, 12)	29.80
	1.82 (br d, 11.5)		2.39 (dd, 3,14)		1.80 (br d, 12)	
7	1.35 (br t, 11.5)	43.43	1.90 (br t, 13.5)	38.19	1.14 (br t, 12)	39.37
8	1.11 (dq, 3,11.5)	24.38	1.41 (dq, 4.5,13.5)	25.09	1.26 (dq, 4,12)	27.97
	1.51 (br d, 11.5)		1.50 (br d, 13.5)		1.41 (br d, 12)	
9	1.38 (dt, 3,11.5)	40.11	1.27 (dt, 13.5,4.5)	35.54	1.41 (br d, 12)	38.13
	1.55 (dt, 11.5,3)		1.82 (dt, 4.5,13.5)		1.49 (dt, 5.5,12)	
10		37.70		41.27		39.07
11	2.05 (m)	33.76	2.05 (m)	34.67	2.00 (m)	33.20
12	0.89 (d, 6.5)	16.72	0.93 (d, 6)	17.64	0.89 (d, 6.5)	16.79
13	2.25 (dd, 8,16)	47.91	2.31 (dd, 9,16)	48.67	2.23 (dd, 8,12)	47.07
	2.44 (dd, 4,16)		2.58 (dd, 4,16)		2.35 (dd, 4,12)	
14		210.56		210.85		210.23
15	2.28 (m)	52.41	2.34 (m)	53.20	2.26 (m)	52.40
16	2.25 (m)	24.59	2.11 (m)	25.53	2.12 (m)	24.52
17	0.92 (d, 6.5)	22.60	0.90 (d, 5)	23.35	0.91 (d, 6.5)	22.52
18	0.93 (d, 6.5)	22.57	0.89 (d, 5)	23.28	0.91 (d, 6.5)	22.59
19	0.85 (s)	17.03	1.02 (s)	24.23	1.18 (s)	21.02
20	1.90 (t, 1.5)	21.84	2.05 (d, 1.5)	22.91	1.98 (d, 1.5)	19.79
OOH			10.29 (s)		8.78 (s)	

TABLE 1. 1 H (δ , J in Hz) and 13 C (δ) NMR Data for Compounds 1-3

References and Notes

- Antitumor Agents 182. For part 181 see: Xia, Y.; Yang, Z. Y.; Xia, P.; Bastow, K. F.; Tachibana, Y.; Kuo, S. C.; Hamel, E.; Hackl, T.; Lee, K. H. J. Med. Chem. 1998, 51, 1155.
- 2. (a) Kashiwada, Y.; Fujioka, T.; Chang, J. J.; Chen I. S.; Mihashi, K.; Lee, K.-H. Bioorg. Med. Chem. Lett. 1992. 2, 395. (b) Idem. J. Org. Chem. 1992. 57, 6946. (c) Fujioka, T.; Sakurai, A.; Mihashi, K.; Kashiwada, Y.; Chen I. S.; Lee, K.-H. Chem. Pharm. Bull. 1997, 45, 68. (d) Idem, ibid, 1997, 45, 202.
- 3. $\left[\alpha\right]_{0}^{20} -50.6^{\circ}$ (c 0.13, acetone). $C_{20}H_{33}O_{2}$ [(M + H)⁺ m/z 305.2478, calcd 305.2480].
- 4. Hikino, H.; Aota, K.; Takemoto, T. Tetrahedron 1971, 27, 4831.
- 5. Babidge, P. J.; Massy-Westropp, R. A. Aust. J. Chem. 1984, 37, 629.
- 6. Mp: 122-123 °C, $[\alpha]_D^{20}-52.8$ ° (c 0.06, acetone). $C_{20}H_{33}O_4$ [(M + H)⁺ m/z 337.2390, calcd 337.2378].
- 7. (a) Ahmad, V. U.; Fizza, K. Liebigs Ann. Chem. 1987, 643. (b) Bohlmann, F.; Wallmeyer, M.; Jakupovic, J.; Gerke, T.; King, R. M.; Robinson, H. Phytochemistry 1985, 24, 505.
- 8. Ahmad, V. U.; Fizza, K.; Amber, A. R. J. Nat. Prod. 1989, 52, 861
- 9. Mp: 129-133 °C, $[\alpha]_D^{20}-69.4$ ° (c 0.17, acetone). $C_{20}H_{33}O_4$ [(M + H)⁺ m/z 337.2386, calcd 337.2378].
- 10. The in vitro cytotoxicity assay was carried out using the National Cancer Institute protocol. Details of the assay procedure have been reported.11
- 11. Boyd, M. B.; Paul, K. D. Drug. Development Res. 1995, 34, 91.

Measured at 400 MHz (¹H) and 100 MHz (¹³C) in CDCl₃
Measured at 500 MHz (¹H) and 125 MHz (¹³C) in acetone-d₆